

Recommended Methods of Euthanasia: *Rabbits*

Species	Method	Description
Rabbits	Inhalant anesthetic overdose followed by bilateral thoracotomy.	Using a precision vaporizer with induction chamber and waste gas scavenger, (<i>indicate the gas anesthetic</i>) will be administered slowly up to [<i>indicate: > 4.5 % (for Isoflurane) or > 6.5 % (for Sevoflurane)</i>] in oxygen and continued until respiratory arrest occurs for > 60 seconds. The chamber is flushed with oxygen only, the animal is removed and bilateral thoracotomy is performed to assure euthanasia.
	Inhalant anesthetic overdose followed by decapitation.	Using a precision vaporizer with induction chamber and waste gas scavenger, (<i>indicate the gas anesthetic</i>) will be administered slowly up to [<i>indicate: > 4.5 % (for Isoflurane) or > 6.5 % (for Sevoflurane)</i>] in oxygen and continued until respiratory arrest occurs for > 60 seconds. The chamber is flushed with oxygen only, the animal is removed and decapitated with (<i>indicate equipment used</i>) to assure euthanasia.
	Inhalant anesthetic overdose followed by exsanguination.	Using a precision vaporizer with induction chamber and waste gas scavenger, (<i>indicate the gas anesthetic</i>) will be administered slowly up to [<i>indicate: > 4.5 % (for Isoflurane) or > 6.5 % (for Sevoflurane)</i>] in oxygen and continued until respiratory arrest occurs for > 60 seconds. The chamber is flushed with oxygen only, the animal is removed and rapid exsanguination is performed by (<i>indicate method or vascular incision points</i>) to assure euthanasia.
	Inhalant anesthetic overdose followed by major organ harvest.	Using a precision vaporizer with induction chamber and waste gas scavenger, (<i>indicate the gas anesthetic</i>) will be administered slowly up to [<i>indicate: > 4.5 % (for Isoflurane) or > 6.5 % (for Sevoflurane)</i>] in oxygen and continued until respiratory arrest occurs for > 60 seconds. The chamber is flushed with oxygen only, the animal is removed and rapid removal of (<i>indicate tissues / organs</i>) is performed to assure euthanasia.
	Vital perfusion under injectable anesthesia	(<i>Indicate drug, dose in mg/kg, route & gauge needle</i>) will be used to induce anesthesia. Surgical depth of anesthesia will be verified by lack of response to (<i>indicate stimulus</i>) stimulus. Vital perfusion will be performed using (<i>indicate name</i>) perfusate injected into the (<i>define point of vascular access or blood egress site</i>). Perfusion will be performed in a chemical fume hood if required by EHS. Perfusate waste will be disposed of by (<i>indicate method of disposal</i>).
	Decapitation by guillotine under sedation or anesthesia	Post induction of anesthesia or sedation using (<i>indicate drug, dose in mg/kg, route & gauge needle</i>) the animal's head is placed to the level of the cervical vertebrae in a commercial guillotine and the guillotine is activated. <i>NOTE: Scientific Justification is REQUIRED for the use of this method without sedation / anesthesia.</i>
	Injectable anesthetic overdose (Pentobarbital)	Administration of ≥ 100 mg/kg of Pentobarbital (<i>state manufacturer</i>) intravascular (IV), intraperitoneal (IP), or intracardiac (IC). Monitor animal until lack of heart beat is noted for > 60 seconds prior to tissue harvest or carcass disposal. <i>NOTE: If utilizing the IC route of administration, indicate / describe details used to produce a surgical plane of anesthesia prior to injection with Pentobarbital.</i>